

Keeping a lab notebook

OITE Bootcamp 2011
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Why keep a lab notebook

- Keep a record of experiments and experimental setup
 - Written records are useful in many settings besides wetlabs.
- To be able to accurately reproduce experiments months or years later
- Allow others to reproduce your experiments
- Aid in trouble-shooting failed experiments
- Primary source for writing the methods for a scientific poster or paper

Types of notebooks: Bound

- Pages are bound or sewn together
- Advantages
 - Portable
 - Inexpensive
 - Pages won't get lost or out of order
 - Can tape in data sheets
- Disadvantages
 - Not easily searchable
 - Can't reorder pages
 - Difficult to copy
 - Requires legible hand-writing



Types of notebooks: Loose-leaf

- Three-ring binder
- Advantages
 - Portable
 - Can insert other experimental data sheets (computer printouts)
- Disadvantages
 - Not easily searchable
 - Requires legible hand-writing
 - Pages may fall out or get out of order

Types of notebooks: Electronic

- Computer program or computer record
 - Advantages
 - Searchable
 - Can be easily backed up and duplicated
 - Can be linked with any electronic data
 - Disadvantages
 - Requires a laptop for portability
 - Need to learn new computer software
 - Theft, damage or file corruption
- Many labs already have standard notebooks, it is best to follow the lab standard

Types of Pens

- Make sure the lab pen you use is fairly resistant to spills
- Do not use pencil
- Avoid sharpies- they can easily bleed through pages, making multiple entries illegible

Pen	Abuse treatment				
	Control	Water	Methanol	Ethanol	Acetone
Bic Accountant fine point (red)	12.3	12.3	12.3	12.3	12.3
Bic Accountant fine pt (black)	12.3	12.3	12.3	12.3	12.3
Bic Round Stic med (black)	12.3	12.3	12.3	12.3	12.3
Cross fountain pen (blue/black)	12.3	12.3	12.3	12.3	12.3
Dixon Ticonderoga 138B-2 soft pencil	12.3	12.3	12.3	12.3	12.3
Pentel Hybrid Gel Roller (black)	12.3	12.3	12.3	12.3	12.3
Pilot G-2 07 (black)	12.3	12.3	12.3	12.3	12.3
Sakura Gelly Roll fine (black)	12.3	12.3	12.3	12.3	12.3
Sakura Gelly Roll fine (blue)	12.3	12.3	12.3	12.3	12.3
Sakura Gelly Roll XPGB (blue)	12.3	12.3	12.3	12.3	12.3
Sanford Uni-Ball Gel RT Med (black)	12.3	12.3	12.3	12.3	12.3
Sanford Uni-Ball Vision fine (black)	12.3	12.3	12.3	12.3	12.3
Sanford Uni-Ball Vision fine (blue)	12.3	12.3	12.3	12.3	12.3
Sanford Uni-Gel RT fine (blue)	12.3	12.3	12.3	12.3	12.3

Image from <http://www.swarthmore.edu/NatSci/cpurris1/notebookadvice.htm>

Front section of notebook

- Your name, lab, project
- Any abbreviations and special notes
 - Sample labeling conventions
- Table of contents
 - Date
 - What experiment was completed
 - Page numbers

Notebook front section example

My Summer intern lab notebook

Name: _____
 Lab: _____
 Project: _____

abbreviations: FCX (frontal cortex), WB (western blot)

Table of contents

Date	Experiment	pg #
7/1/11	WB to examine protein levels in FCX	20-4
7/3/11	antibody testing	5-7
7/4/11	HeLa cell transfection	8
7/10/11	purifying nuclei from cells	9-11

Notebook entry organization

- **Headings:** Date, title of experiment
- **Body:** All steps of experimental protocol (including deviations from prescribed protocol) Buffer compositions, any observations
- **Closing:** Results (include pictures if possible)
- If your notebook doesn't have numbered pages, number them

Notebook entries are not unlike cooking recipes

Notebook Example

8/12/11 macaroni & cheese w/ chocolate milk
 Objective: make lunch

Recipe for chocolate milk (CM):

milk 1 cup
 cocoa 2 tbsps
 - stir until cocoa changes from white to brown

Macaroni:

4 cups of water in pot on stove & turned burner to high heat
 - this will make water boil
 - Once water is boiling, add macaroni
 - make sure to remove cheese packet
 - let macaroni cook for 6 minutes
 - stir every 3 minutes
 - Used colander to drain water
 - Transferred macaroni from colander to pot
 - Turned off burner
 - add cheese packet, 1 tbs butter, & CM to macaroni
 - mix with wooden spoon until smooth
 - Serve in bowl with CM

"Best Practices" Lab notebook entry

Date and experiment title	Objective or hypothesis
Aug 12, 2011 Nuclear purification Objective: Purify nuclei from transfected cells to examine protein localization	
Very detailed descriptions	Description of buffer Molarity, and what was weighed Manufacture and catalog # of reagents
Homogenization Buffer (HB) 0.32M Sucrose (Sigma #0012) 11g 0.15M MgCl ₂ (Sigma #3412) 1.92g Qro 100 ml w/ dH ₂ O	
Explanation of each step	Always include centrifuge conditions Always include measurements
<ul style="list-style-type: none"> • Cellated 90% confluent cells from 100mm² plate • centrifuged 100g, 5 minutes, 4°C - this step pellets cells in dense medium • resuspended cell pellet in 10ml HB • homogenized cells w/ glass homogenizer and Teflon tissue grinder - this step breaks open cells while keeping nuclei intact • pelleted nuclei by centrifuging 12000g, 10 minutes, 4°C • removed & discarded supernatant • stored nuclei pellet in -80°C freezer 	
	Labeled storage space

Make use of prewritten protocols

- Plan each experiment out before beginning
- Can tape typed protocols in notebook as long as there is room for edits

Nuclear Purification Protocol August 12, 2011	
Homogenization Buffer	weighed out
0.32M Sucrose (Sigma)	11g
0.15M MgCl ₂ (Sigma)	1.92g
added H ₂ O to 50ml final volume	
1. Collect cells and remove media - 100mm plate 90% confluent	
Centrifuge 1000g, 5min, 4°C to pellet cells - remove supernatant	
2. Resuspend cells in cold, freshly made HB buffer	
- used 2ml of buffer	
3. Homogenize with glass homogenizer and Teflon pestle	
- breaks open cells	
4. Pellet nuclei (20,000g or 14,000rpm, 10 minutes, 4°C)	
5. Discard supernatant	
Stored samples in -80°C freezer, bottom shelf	

